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Michael P. Maher

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EXAMINER

PAK, MICHAEL D

ART UNIT

PAPER NUMBER

1646

DATE MAILED: 05/16/2006

Please find below and/or attached an Office communication concerning this application or proceeding.

DETAILED ACTION

Response to Amendment

1. The claims and response filed 28 October 2005 has been entered.
2. The text of those sections of Title 35, U.S. Code not included in this action can be found in a prior Office action.
3. Applicant's arguments filed 28 October 2005, have been fully considered but they are not found persuasive.

Claim Rejections - 35 USC § 103

4. Claims 1-8, 10-29, 49 and 50 are rejected under 35 U.S.C. 103(a) as being unpatentable over Gonzalez et al. (1995) in view of Reiner et al. (1995).

The reason for the rejection has been set forth previously.

Gonzalez et al. teaches a method for achieving fast ratiometric voltage-sensitive fluorescence changes in single cells using fluorescence resonance energy transfer. The mechanism is based on hydrophobic fluorescent anions that rapidly redistribute from one face of the plasma membrane to the other according to the Nernst equation (Gonzalez, page 1272). In this method L-M(TK-) fibroblasts were loaded with DIBAC(3), a fluorescent dye to monitor membrane potential transients, and coated with fluorescein-labeled wheat germ agglutinin (Ibid, page 1273), this pair serves as the donor-acceptor pair for the energy transfer. The L-M(TK-) cells have low background

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currents (Ibid, page 1275). The method teaches the measurement of fluorescent changes of the DISBAC(3) in response to voltage changes (Ibid, page 1276, figure 4). The change in the transmembrane potential is measured without the use of the patch clamp technique. In this Figure, the voltage steps are applied with the patch clamp technique, while the output is measured by monitoring the fluorescence. The electric field would not vary over the area of observation, which in this case is a single cell. Gonzalez further teaches monitoring the fluorescence intensity changes, indicative of membrane potential, in response to square wave step depolarizations from the -70mV holding potential to 40, 80, 120 and 160 mV (Ibid, page 1277, Figure 6). Again, the voltage steps are applied with the patch clamp technique, while the indicia of membrane potential is the fluorescence intensity. The step potentials are applied for 500 milliseconds. Gonzalez et al. further teaches the practice of this method in neonatal cardiac myocytes, which comprise voltage gated ion channels, which are activated upon depolarization (Ibid, page 1278, figure 8). Gonzalez et al. does not teach characterizing the effect of a compound on ion channel activity of a compound with this method.

Renier et al. teaches a method to evaluate expression of functional CFTR. The technique uses the potential-sensitive probe DISBAC2(3), by single-cell fluorescence imaging. The DISBAC(3) method was first validated on the mouse mammary tumor cell line C127, stably expressing wild-type CFTR (Renier, page 1278, Figure 1). Activation of protein kinase A by the cAMP-permeable analogue 8-Br-cAMP induced cell membrane depolarization consistent with expression of wild-type CFTR. The effect of 8-Br-cAMP on A549 cells transfected with adenovirus encoding CFTR was then

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measured (Ibid, page 1279, Figure 2). Therefore, it would have been obvious to one of ordinary skill in the art at the time the invention was made to practice a method of characterizing the effect of a compound on ion channel function by exposing a cell expressing the ion channel to alterations in the electric field and measuring the effect on the membrane potential with fluorescent dyes. The motivation is provided in the Renier reference which teaches that the DISBAC(3) method is quick, simple, and reproducible, and does not require invasive cell loading procedures (Ibid, page 1275). The expectation of success is provided in the Gonzalez reference which teaches that voltage indicators based on FRET may already be practically useful and that modest, rationally attainable improvements in sensitivity and speed could make them superior for many biological applications.

5. No claims allowed.

6. Any inquiry concerning this communication or earlier communications from the examiner should be directed to Michael Pak, whose telephone number is (571) 272-0879. The examiner can normally be reached on Monday through Friday from 8:30 AM to 2:00 PM.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Gary Nickol, can be reached on (571) 272-0835.

The fax phone number for the organization where this application or proceeding is assigned is 703-872-9306. Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or

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Michael Pak
Primary Patent Examiner
Art Unit 1646
13 May 2006